

Body mass scaling of passive oxygen diffusion in endotherms and ectotherms

James F. Gillooly^{a,1}, Juan Pablo Gomez^{a,b}, Evgeny V. Mavrodiev^b, Yue Rong^c, and Eric S. McLaMORE^c

^aDepartment of Biology, University of Florida, Gainesville, FL 32611; ^bFlorida Museum of Natural History, University of Florida, Gainesville, FL 32611; and ^cDepartment of Agricultural and Biological Engineering, University of Florida, Gainesville, FL 32611

Edited by Mary I. O'Connor, University of British Columbia, Vancouver, BC, Canada, and accepted by the Editorial Board March 30, 2016 (received for review October 2, 2015)

The area and thickness of respiratory surfaces, and the constraints they impose on passive oxygen diffusion, have been linked to differences in oxygen consumption rates and/or aerobic activity levels in vertebrates. However, it remains unclear how respiratory surfaces and associated diffusion rates vary with body mass across vertebrates, particularly in relation to the body mass scaling of oxygen consumption rates. Here we address these issues by first quantifying the body mass dependence of respiratory surface area and respiratory barrier thickness for a diversity of endotherms (birds and mammals) and ectotherms (fishes, amphibians, and reptiles). Based on these findings, we then use Fick's law to predict the body mass scaling of oxygen diffusion for each group. Finally, we compare the predicted body mass dependence of oxygen diffusion to that of oxygen consumption in endotherms and ectotherms. We find that the slopes and intercepts of the relationships describing the body mass dependence of passive oxygen diffusion in these two groups are statistically indistinguishable from those describing the body mass dependence of oxygen consumption. Thus, the area and thickness of respiratory surfaces combine to match oxygen diffusion capacity to oxygen consumption rates in both air- and water-breathing vertebrates. In particular, the substantially lower oxygen consumption rates of ectotherms of a given body mass relative to those of endotherms correspond to differences in oxygen diffusion capacity. These results provide insights into the long-standing effort to understand the structural attributes of organisms that underlie the body mass scaling of oxygen consumption.

allometry | metabolic theory | respiration rate | metabolism | oxygen consumption

Since the classic debates of Krogh and Bohr (1), the relationship of passive oxygen diffusion to oxygen consumption has remained the focus of considerable research (2). Still, we have much to learn about how oxygen diffusion across respiratory surfaces relates to whole-organism oxygen consumption (3). In particular, we have much to learn about how the structural constraints on oxygen diffusion capacity, namely the thickness and area of respiratory surfaces, relate to the body mass scaling of oxygen consumption in vertebrates—a pattern that reflects the scaling of species' energy use (3).

Studies on the body mass dependence of oxygen diffusion have been undertaken over the last century in all major classes of vertebrates, typically using respiratory surface area as a metric of diffusion capacity (2, 3). Less commonly, studies have also considered potentially relevant changes in respiratory barrier thickness with mass (4). In many cases, these studies have concluded that the body mass scaling of oxygen diffusion relative to that of oxygen consumption differs between endotherms and ectotherms (3, 5, 6). This has led to very different conclusions regarding the structure and function of respiratory systems among classes of vertebrates, as explained below.

Among endotherms, particularly mammals, researchers have concluded that a mismatch exists between the body mass scaling of oxygen diffusion and oxygen consumption (6). Oxygen diffusion capacity is thought to scale linearly with body mass (or nearly so) in these groups, whereas oxygen consumption is thought to increase less than proportionally with mass (3, 6–8). Thus, when

these relationships with body mass are expressed as power law functions of the form $Y = M^b$, the scaling exponent “b” is about 1 for diffusion, but about 2/3 to 8/10 for consumption (9–12). This apparent mismatch has confounded modeling efforts to explain the body mass dependence of oxygen consumption rates in mammals and other vertebrates (e.g., refs. 8 and 13). It has also been used as evidence that the structure of respiratory systems has evolved to optimize performance at maximum rather than minimum rates of oxygen consumption. This is because maximum consumption seems to scale more steeply with body mass, like respiratory surface area (14). More generally, this apparent mismatch in the scaling of oxygen diffusion and consumption has been used to argue against the evolutionary optimization principle of symmorphosis (15) in complex biological systems such as the respiratory system (16, 17).

In contrast, among some classes of ectotherms, previous work has concluded that the body mass scaling of oxygen diffusion capacity roughly matches that of oxygen consumption (3, 5, 18). In fishes, for example, the relationships of both oxygen consumption and respiratory surface area are thought to scale to the 0.6–0.8 power of body mass (5). These observations have led to the argument that passive oxygen diffusion is a rate-limiting step in oxygen consumption, and that the size or thickness of respiratory surface areas arise from evolutionary tradeoffs in animal design (5, 19).

Within studies of both endotherms and ectotherms, researchers have noted that species with greater oxygen diffusion capacity and larger respiratory surface areas tend to show higher levels of aerobic activity or “athleticism” (19–22). Between these two groups, however, it remains unclear to what extent respiratory surface area and/or other physiological differences (e.g., mitochondrial density)

Significance

Biologists have long sought to understand how differences in the structure of vertebrate respiratory systems are related to differences in oxygen consumption rates associated with metabolism. In particular, they have sought to understand how structural constraints on passive oxygen diffusion through gills or lungs are related to the nonlinear body mass scaling of oxygen consumption rates. Here we show that the body mass dependence of oxygen diffusion capacity, which is governed by the area and thickness of respiratory surfaces, matches the body mass dependence of respiration in diverse endothermic (birds and mammals) and ectothermic (reptiles, amphibians, and fishes) vertebrates. These results provide a step toward a more complete understanding of the structural differences that underlie changes in oxygen consumption rates with body size.

Author contributions: J.F.G. designed research; J.F.G., J.P.G., E.V.M., Y.R., and E.S.M. performed research; J.F.G., J.P.G., and E.V.M. analyzed data; and J.F.G. and J.P.G. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission. M.I.O. is a guest editor invited by the Editorial Board.

¹To whom correspondence should be addressed. Email: gillooly@ufl.edu.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1519617113/-DCSupplemental.

may correspond to the substantially lower oxygen consumption rates of ectotherms vs. endotherms at a given body mass (22). General quantitative relationships between the structural constraints on oxygen diffusion and oxygen consumption have not been established across vertebrates.

Thus, here we quantitatively examine the body mass dependence of the two primary structural constraints on oxygen diffusion capacity in ectotherms and endotherms, namely respiratory surface area and respiratory barrier thickness (air–blood barrier in lungs and water–blood barrier in gills). Based on these results, we then predict the overall rate of oxygen diffusion as a function of body mass in ectotherms and endotherms using Fick’s law of diffusion (23). Finally, we compare the predicted scaling relationships of oxygen diffusion to observed relationships for oxygen consumption in these two groups. Analyses of the thickness and area of respiratory surfaces, as well as oxygen consumption rates, include species from all major classes of vertebrates (birds, mammals, fishes, amphibians, and reptiles).

Model Development

Following from Fick’s law of diffusion (23), oxygen diffusion can be modeled as a passive, two-dimensional process that is structurally constrained by two factors: the total respiratory surface area (RSA, in square centimeters) over which gas exchange occurs and the respiratory barrier thickness of that barrier (RBT, in centimeters). In this formulation, diffusion rate is the product of the oxygen flux per unit respiratory surface area $\left(\frac{\Delta pO_2 * K}{RBT}\right)$ multiplied by the total RSA such that

$$\frac{\text{mL O}_2}{\text{hour}} = \left(\frac{\Delta pO_2 * K}{RBT}\right) (\text{RSA}), \quad [1]$$

where ΔpO_2 is the partial pressure gradient of oxygen (mm Hg) and K is Krogh’s diffusion constant ($\text{cm}^2 \cdot \text{h}^{-1} \cdot \text{mL O}_2 \cdot \text{cm}^{-3} \cdot \text{tissue} \cdot \text{mm Hg}^{-1}$). Both K and ΔpO_2 are assumed here to be independent of body mass, consistent with previous work (6, 24). Thus, the body mass dependence in Eq. 1 arises from the body mass dependence of RSA and RBT, which can be described by power laws of the form $\text{RSA} = aM^\alpha$ and $\text{RBT} = bM^\beta$, where M is body mass (grams) (4, 25). Substituting these equations for RSA and RBT into Eq. 1 yields a general expression for the body mass dependence of diffusion rate:

$$\frac{\text{mL O}_2}{\text{hour}} = \left(\frac{a}{b}\right) (\Delta pO_2 * K) (M^{\beta-\alpha}), \quad [2]$$

where the overall mass dependence is determined by the difference between the scaling of RSA and RBT ($M^{\beta-\alpha}$). Note that although Eq. 2 describes a passive process, active processes to enhance oxygen delivery may contribute to diffusion through effects on ΔpO_2 (26).

Eq. 2 can be parameterized to yield quantitative predictions for oxygen diffusion rate as a function of body mass at the whole organism level. We do so below for endotherms and ectotherms based on reported values of ΔpO_2 and K , and the empirical relationships of RSA and RBT to body mass presented here.

Methods

RBT and RSA. Published data were compiled on RBT and RSA for all major classes of vertebrates (birds, mammals, amphibians, fishes, and reptiles), which includes both air-breathing species with lungs and water-breathing species with gills. Data broadly represent the taxonomic diversity, body sizes, body temperatures, and aerobic activity levels that occur in each class (SI Appendix, Appendix 1). As such, these data include species from all major ecosystems (marine, freshwater, and terrestrial), using all major forms of locomotion (swimming, flying, and walking) and exhibiting a broad range of life histories. However, efforts were made to restrict analyses to adult or subadult individuals because respiratory features may change through ontogeny (4, 27).

Body mass estimates corresponding to RBT and RSA values were taken from the original studies or, if unavailable, from other published sources (SI Appendix, Appendix 1).

Standard morphometric measures of RBT and RSA were used in all analyses. RBT is described using the harmonic mean distance, which is considered the most relevant measure with respect to gas exchange (4). For air-breathing species, this represents the air–blood barrier distance and for water breathers the water–blood barrier distance. Measures reported as arithmetic mean thickness (AMT) were converted to harmonic mean thickness (HMT) based on the published equation $\text{HMT} = 2/3 \times \text{AMT}$ for species other than birds (28). Respiratory surface areas were measured on whole lungs or gills, upon removal and fixation, following standard procedures (25).

Quantifying RBT and RSA in amphibians presents unique challenges because oxygen diffusion occurs through the lungs, mouth cavity, and skin to varying degrees (29). Thus, it was necessary to approximate RBT and RSA in amphibians in a manner that differed from other species considered here in two respects. First, RSA in amphibians was based on the total respiratory capillary surface areas in the lungs, mouth cavity, and skin. For anuran amphibians, this estimate was based on the total length of respiratory capillaries for each species, and the relationship between total capillary length and total capillary area described for caudate amphibians (30). Second, for RBT, a single mean value from lungs was used for all caudates ($n = 4$) (4), and species-specific estimates from lungs were used for all anurans. Given these approximations, we present statistics for ectotherms both with and without amphibians.

ΔpO_2 and Diffusion Rate. Data were compiled on the partial pressure difference of oxygen across the respiratory barrier for endotherms and ectotherms at rest (SI Appendix, Appendix 2). For species with lungs, ΔpO_2 estimates represent the partial pressure difference between the air sacs of lungs on one side of the barrier and blood vessels on the other. This equates to the alveolar-to-arterial pO_2 difference in mammals or, for vertebrate classes lacking alveoli, the difference between blood pressure and the partial pressure of air in comparable lung structures [e.g., parabronchi in birds; see ref. 31]. For species with gills, ΔpO_2 represents the difference between the partial pressure of oxygen in inhaled water and that in afferent blood vessels.

The Krogh’s diffusion constant (K ; $\text{cm}^2 \cdot \text{h}^{-1} \cdot \text{mL O}_2 \cdot \text{cm}^{-3} \cdot \text{tissue} \cdot \text{mm Hg}^{-1}$) used in determining overall oxygen diffusion rates for both endotherms and ectotherms represents the product of the rate of diffusion (cm^2/h) multiplied by the solubility of oxygen in tissue ($\text{mL O}_2 \cdot \text{cm}^{-3} \cdot \text{tissue} \cdot \text{mm Hg}^{-1}$) (32). The value used here was based on in vivo measures of oxygen diffusion using phosphorescence quenching microscopy (32). Although this estimate is considerably lower than Krogh’s original estimate (33), it is within two- to threefold of many current estimates from various tissues—particularly those using in vivo methods (34–46). The diffusion constant was adjusted for temperature to 38 °C in our analysis of endotherms and to 25 °C for ectotherms, based on a reported Q_{10} of 1.1 (47).

Oxygen Consumption Rates. For oxygen consumption of endotherms at rest, we relied primarily on two recent compilations of data (48, 49). For oxygen consumption rates of ectotherms at rest, we relied exclusively on the compilation of (48) (SI Appendix, Appendix 3). Because this dataset often included multiple points for a single species, we included only the individual(s) at the largest body mass and highest constant temperature. For both endotherms and ectotherms, most oxygen consumption measures were taken from subadult or adult individuals.

Statistical Analyses. Statistical models were fit to natural log-transformed data using both ordinary least squares (OLS) and phylogenetic generalized least squares [PGLS (50); regression with R version 3.0 (51)] (Table 1). For PGLS, because no well-resolved phylogeny exists for all vertebrates, we constructed our own phylogenetic supertree based on recent phylogenies of each vertebrate class [fishes (52), mammals (53), amphibians (54), birds (55), and reptiles (56–59); see also ref. 60]. The tree was constructed using the matrix representation with parsimony (MRP) approach (61, 62) and the Baum–Ragan coding procedure (63). Execution of the final MRP matrix was performed in TNT under the defaults for “traditional search” (64), and conversions between tree formats were performed in Mesquite 2.75 (65). This approach to supertree construction is widely used and has been shown to be effective (66).

For the PGLS analyses, we treated polytomies as soft (67) and assumed equal branch lengths given the uncertainty in divergence times within and across classes of vertebrates (68). The expected covariance among species was estimated using the *corBrownian* function in the APE package for R (69), which assumes a Brownian motion model of trait evolution (70, 71). In

Table 1. Summary of OLS and PGLS regression analyses for the body mass dependence of RBT, RSA, and oxygen consumption rates in endothermic (birds and mammals) and ectothermic (fishes, amphibians, and reptiles) vertebrates

Models	N	df	I	95% CI	P	M	95% CI	P	R ²
RBT, μm									
Endotherms									
PGLS (Fig. 1B)	56	54	0.13	(0.09,0.21)	**	0.1	(0.06,0.14)	**	0.32
OLS	59	57	0.13	(0.1, 0.17)	**	0.11	(0.08,0.15)	**	0.43
Ectotherms									
PGLS (Fig. 1B)	33	31	1.85	(1.05,3.25)	0.04	-0.04	(-0.11,0.03)	0.27	0.03
PGLS-NoA	18	16	2.97	(1.15,7.69)	0.04	-0.11	(-0.23,0.01)	0.1	0.12
OLS	44	42	1.91	(1.27, 2.91)	**	-0.04	(-0.12,0.04)	0.08	0.02
RSA, cm^2									
Endotherms									
PGLS(Fig. 1A)	48	46	58.55	(31.5,109.9)	**	0.89	(0.83,0.95)	**	0.98
OLS	51	49	57.97	(45.6,74.44)	**	0.9	(0.86,0.93)	**	0.98
Ectotherms									
PGLS(Fig. 1A)	24	22	6.88	(2.69,17.63)	**	0.78	(0.65,0.9)	**	0.81
PGLS-NoA	12	10	3.53	(0.81,13.59)	0.12	0.85	(0.64,1.06)	**	0.91
OLS	33	31	8.93	(5.15,15.33)	**	0.76	(0.64,0.87)	**	0.85
O₂ consumption, mL O₂/h									
Endotherms									
PGLS (Fig. 2A)	580	578	3.28	(1.95,5.58)	**	0.75	(0.73,0.78)	**	0.95
OLS	580	578	4.09	(3.78,4.39)	**	0.7	(0.69,0.71)	**	0.95
Ectotherms									
PGLS (Fig. 2B)	249	247	0.13	(0.02,0.63)	**	0.84	(0.8,0.89)	**	0.80
OLS	249	247	0.10	(0.08,0.13)	**	0.9	(0.85,0.95)	**	0.82
O₂ diffusion									
Endotherms									
Ectotherms									

The character "I" represents the intercept of the scaling relationship and "M" represents the scaling exponent. *P* values of *P* < 0.001 are noted with asterisks. Statistics listed as "NoA" represent those without amphibians (Methods). Data and sources are listed in [SI Appendix, Appendixes 1 and 3](#).

Table 1, we present the results of both OLS and PGLS analyses because some subset of species were not present in the phylogeny.

Results

Analyses reveal stark differences in the relationships of RSA and RBT to body mass between ectotherms and endotherms, after accounting for any influence of evolutionary relatedness among species. RSA shows strong, positive relationships with mass in both groups but increases more steeply with mass in endotherms

than in ectotherms ($\text{RSA} \propto M^{0.89}$ vs. $\text{RSA} \propto M^{0.78}$; Fig. 1A and Table 1). In contrast, RBT increases modestly with body mass in endotherms and shows no significant increase with mass in ectotherms ($\text{RBT} \propto M^{0.1}$ in endotherms vs. $\text{RBT} \propto M^{-0.04}$ in ectotherms; Fig. 1B and Table 1). For a given body mass, RBT is about 13-fold higher, and RSA is about 8- to 10-fold lower, in ectotherms than in endotherms (Fig. 1, Table 1, and [SI Appendix, Appendix 1](#)). Body mass also explained substantially more variation in RSA than RBT across species (Table 1).

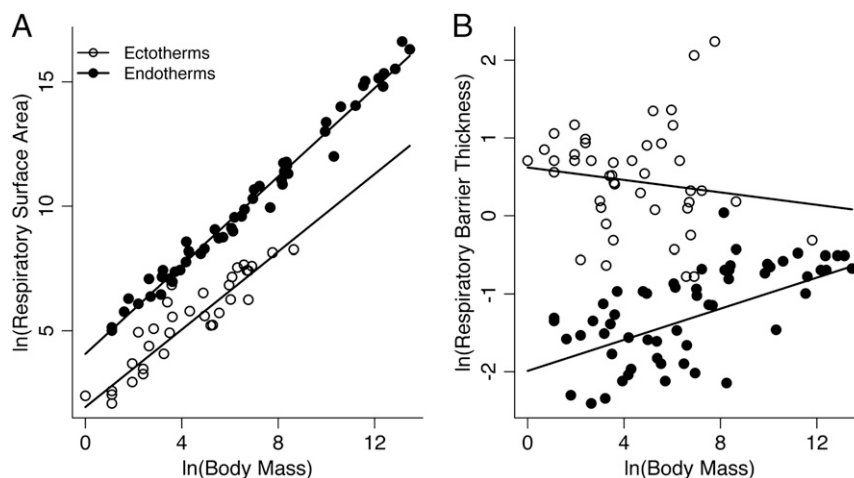


Fig. 1. The relationships of (A) RSA (square centimeters) and (B) RBT (micrometers) to body mass (grams) in endotherms (birds and mammals) and ectotherms (reptiles, fishes, and amphibians). Statistics for the lines, fit using PGLS regression, are given in Table 1. Data and sources are listed in [SI Appendix, Appendix 1](#).

The overall body mass dependence of oxygen diffusion rate in endotherms and ectotherms is estimated based on the relationships for RSA and RBT shown in Fig. 1. For endotherms at rest, the relationship of oxygen diffusion to body mass is estimated to be $\text{mL O}_2/\text{h} = 4.78 \cdot \text{M}^{0.79}$, based on a mean ΔpO_2 value of 4.28 ($n = 11$), and $K = 2.48 \times 10^{-7} \text{ cm}^2 \cdot \text{h}^{-1} \cdot \text{mL O}_2 \cdot \text{cm}^{-3} \text{ tissue} \cdot \text{mm Hg}^{-1}$ at 38 °C (*Methods*). For ectotherms at rest, the relationship is estimated to be $\text{mL O}_2/\text{h} = 0.15 \cdot \text{M}^{0.82}$, based on a mean ΔpO_2 value of 19.03 ($n = 13$, *SI Appendix, Appendix 2*), and $K = 2.19 \times 10^{-7} \text{ cm}^2 \cdot \text{h}^{-1} \cdot \text{mL O}_2 \cdot \text{cm}^{-3} \text{ tissue} \cdot \text{mm Hg}^{-1}$ at 25 °C. These relationships for the body mass dependence of oxygen diffusion are statistically indistinguishable from those of oxygen consumption in endotherms and ectotherms, based on the 95% confidence intervals (CI) (Table 1). PGLS regression analysis of the body mass dependence of oxygen consumption rates for endotherms at rest yields a fitted line ($\text{mL O}_2/\text{h} = 3.28 \cdot \text{M}^{0.75}$, $n = 580$) that is similar in both slope and intercept to that predicted for diffusion ($\text{mL O}_2/\text{h} = 4.78 \cdot \text{M}^{0.79}$; Fig. 2A and Table 1). The relationship of oxygen consumption rate to body mass in ectotherms also yields a fitted line ($\text{mL O}_2/\text{h} = 0.13 \cdot \text{M}^{0.84}$, $n = 249$) that closely matches that of diffusion rate ($\text{mL O}_2/\text{h} = 0.15 \cdot \text{M}^{0.82}$; Fig. 2B and Table 1).

Discussion

Our results yield estimates of the body mass dependence of passive oxygen diffusion in endothermic and ectothermic vertebrates using Fick's law (23). The body mass dependence of the flux per unit area ($\propto 1/\text{RBT}$) multiplied by the total respiratory area (RSA) combine to determine the scaling of oxygen diffusion shown in Eq. 1. The slopes (i.e., scaling exponents) of these relationships arise from the body mass scaling of respiratory surface area and, to a lesser extent, the scaling of respiratory barrier thickness. In endotherms, RSA scaled to the 0.89 power of body mass, but diffusion scaled to the 0.79 power given the scaling of RBT with mass. In ectotherms, the scaling of diffusion (slope = 0.82) more closely matched the scaling of respiratory surface area (slope = 0.78) (Table 1). The nearly 15-fold variation in the RBT of ectotherms was significantly greater than that of endotherms and showed a much weaker correlation with body mass (Fig. 2 and Table 1).

Difference in the intercepts of the relationships of diffusion with body mass, whereby diffusion rates were roughly 30-fold lower in ectotherms than in endotherms (Fig. 2), also arose largely from differences in RSA and RBT between groups. However, these differences were offset somewhat by the 4.4-fold higher value

of ΔpO_2 in ectotherms than in endotherms (19.03 mm Hg vs. 4.28 mm Hg; *SI Appendix, Appendix 2*). The weak temperature dependence of Krogh's diffusion constant, K , has only a very minor effect on the observed difference in intercepts.

The estimated scaling of oxygen diffusion, based on the scaling of RSA and RBT, was statistically indistinguishable from that of oxygen consumption for both endotherms and ectotherms—consistent with the concept of symmorphosis (15). Incorporating the body mass dependence of RSA and RBT into Fick's law, along with estimates of K and ΔpO_2 , predicts both the slopes and intercepts of the oxygen consumption relationships shown in Fig. 2—there are no free parameters. In endotherms, previous models of oxygen consumption in mammals have assumed a linear scaling of diffusion capacity based on available data (8, 20, 72), and thus a mismatch between diffusion and consumption. To address this mismatch, these models further assumed ΔpO_2 scales to the $-1/12$ power of body mass without supporting data (24). The scaling of oxygen diffusion (slope = 0.79) we observed in endotherms was consistent with the $3/4$ scaling of resting oxygen consumption rates found here, but also with the most recent estimate of the scaling of maximum oxygen consumption rates in mammals (slope = 0.83; CI: 0.79–0.89) (73). This observation also does not seem consistent with the argument that the structure of respiratory systems has evolved to optimize performance at maximum rather than minimum oxygen consumption rates. Moreover, differences in the intercepts between minimum and maximum oxygen consumption rates could potentially be explained solely based on differences in ΔpO_2 . The three- to sixfold difference between minimum and maximum oxygen consumption (12) seems to correspond with a change in ΔpO_2 of similar magnitude from rest to activity in mammals (74–76). In ectotherms, although the scaling of diffusion and consumption matched, both differed significantly from the previously proposed values of $2/3$ and $3/4$ —consistent with previous work (48). OLS and PGLS models of these and other relationships yielded similar results, although they are not strictly comparable (*Methods*).

However, our results should not be viewed as pointing to structural constraints on oxygen diffusion as the single rate-limiting step of oxygen consumption. Passive diffusion is just the first step in the complex system that governs the acquisition and delivery of oxygen. Moreover, we recognize that our use of Fick's law does not account for the many structural and functional differences among species that may affect diffusion (31). We might expect, however, structures

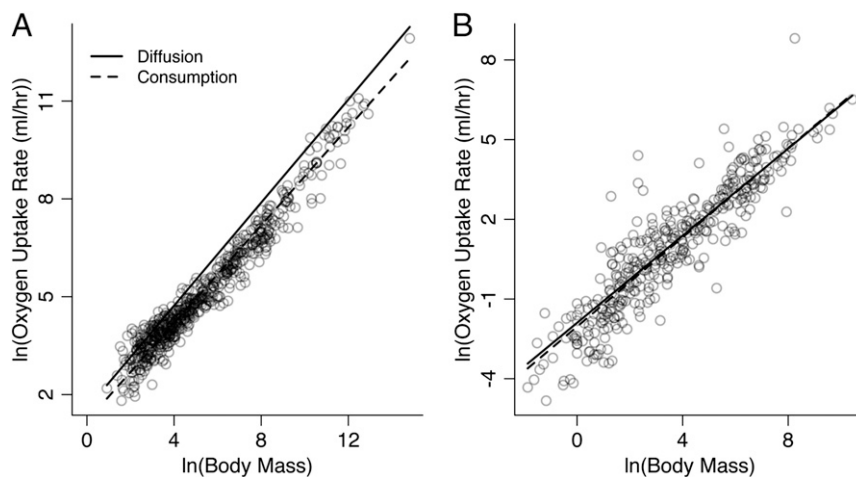


Fig. 2. The observed body mass dependence of oxygen consumption and the predicted body mass dependence of oxygen diffusion for (A) endotherms at rest (birds and mammals; $n = 580$) and (B) ectotherms at rest (reptiles, fishes, and amphibians; $n = 249$). Dashed lines are fitted to the oxygen consumption rate data using PGLS regression (Table 1). Solid lines are the predicted relationships of oxygen diffusion (milliliters of O_2 per hour) to body mass (grams) following Eq. 1. Data and sources are listed in *SI Appendix, Appendixes 1–3*.

relevant to the delivery of oxygen “downstream” of diffusion to show corresponding relationships with mass. For example, we speculate that the size and number of oxygen-carrying red blood cells may be related to the flux per unit area and total area of respiratory surfaces. It is perhaps no coincidence that species with thicker respiratory barriers (e.g., salamanders, lungfish, and tortoises) also tend to have larger cells (77). Passive diffusion may be just one of many colimiting steps on oxygen consumption (78).

Still, the results presented here show how the thickness and area of respiratory surfaces combine to match the body mass dependence of oxygen diffusion to oxygen consumption in endothermic and ectothermic vertebrates. Future work examining the functional tradeoffs associated with differences in RSA and RBT among species may provide insights into the evolutionary forces that have ultimately constrained diffusion rates and perhaps

oxygen consumption rates. For example, perhaps the sizes of RSA and RBT are the result of a tradeoff between maximizing gas exchange and minimizing respiratory water (or ion) loss (79). Respiratory water loss may constitute a significant fraction of total water loss among vertebrates (80), and it seems from our data that species facing greater challenges with respect to water loss (e.g., desert tortoises, lungfish, and salamanders) have relatively small respiratory surface areas and/or relatively thick respiratory barriers. A closer look at diffusion in this way may prove valuable to answering the long-standing question of why oxygen consumption rate, and thus species' energy use (i.e., metabolic rate), scales with body mass.

ACKNOWLEDGMENTS. We thank A. Hein, A. P. Allen, C. Hou, and members of the Network for Ecological Theory Integration working group for helpful comments and discussion.

- Gjedde A (2010) Diffusive insights: On the disagreement of Christian Bohr and August Krogh at the Centennial of the Seven Little Devils. *Adv Physiol Educ* 34(4):174–185.
- Mortola JP (2015) Generalities of gas diffusion applied to the vertebrate blood–gas barrier. *The Vertebrate Blood–Gas Barrier in Health and Disease*, ed Makanya AN (Springer, New York), pp 1–14.
- Hughes GM (1984) Scaling of respiratory areas in relation to oxygen consumption of vertebrates. *Experientia* 40(6):519–524.
- Maina JN, West JB (2005) Thin and strong! The bioengineering dilemma in the structural and functional design of the blood-gas barrier. *Physiol Rev* 85(3):811–844.
- Pauly D (1981) The relationships between gill surface area and growth performance in fish: A generalization of von Bertalanffy's theory of growth. *Meeresforschung* 28(4): 251–282.
- Weibel ER, et al. (1981) Design of the mammalian respiratory system. IX. Functional and structural limits for oxygen flow. *Respir Physiol* 44(1):151–164.
- Lindstedt SL (1984) Pulmonary transit time and diffusing capacity in mammals. *Am J Physiol* 246(3 Pt 2):R384–R388.
- West GB, Brown JH, Enquist BJ (1997) A general model for the origin of allometric scaling laws in biology. *Science* 276(5309):122–126.
- White CR, Kearney MR (2014) Metabolic scaling in animals: Methods, empirical results, and theoretical explanations. *Compr Physiol* 4(1):231–256.
- Agutter PS, Wheatley DN (2004) Metabolic scaling: Consensus or controversy? *Theor Biol Med Model* 1(1):13.
- Price CA, et al. (2012) Testing the metabolic theory of ecology. *Ecol Lett* 15(12): 1465–1474.
- Savage VM, et al. (2004) The predominance of quarter power scaling in biology. *Funct Ecol* 18:257–282.
- Darveau C-A, Suarez RK, Andrews RD, Hochachka PW (2002) Allometric cascade as a unifying principle of body mass effects on metabolism. *Nature* 417(6885):166–170.
- Suarez RK, Darveau C-A, Childress JJ (2004) Metabolic scaling: A many-splendored thing. *Comp Biochem Physiol B Biochem Mol Biol* 139(3):531–541.
- Weibel ER, Taylor CR, Hoppeler H (1991) The concept of symmorphosis: A testable hypothesis of structure-function relationship. *Proc Natl Acad Sci USA* 88(22): 10357–10361.
- Garland T, Huey RB (1987) Testing symmorphosis: Does structure match functional requirements? *Evolution* 41(6):1404–1409.
- Dudley R, Gans C (1991) A critique of symmorphosis and optimality models in physiology. *Physiol Zool* 64:627–637.
- Tenney SM, Tenney JB (1970) Quantitative morphology of cold-blooded lungs: Amphibia and reptilia. *Respir Physiol* 9(2):197–215.
- Palzenberger M, Pohla H (1992) Gill surface area of water-breathing freshwater fish. *Rev Fish Biol Fish* 2(3):187–216.
- Weibel ER, Hoppeler H (2005) Exercise-induced maximal metabolic rate scales with muscle aerobic capacity. *J Exp Biol* 208(Pt 9):1635–1644.
- De Jager S, Dekkers WJ (1974) Relations between gill structure and activity in fish. *Neth J Zool* 25(3):276–308.
- Perry S (2012) Recent advances and trends in the comparative morphometry of vertebrate gas exchange organs. *Vertebrate Gas Exchange: From Environment to Cell*, Advances in Comparative and Environmental Physiology, ed Boutilier RG (Springer, New York), Vol 6, pp 45–71.
- Fick A (1855) Ueber diffusion. *Ann Phys* 170(1):59–86.
- Dawson TH (2005) Modeling of vascular networks. *J Exp Biol* 208(Pt 9):1687–1694.
- Gehr P, et al. (1981) Design of the mammalian respiratory system. V. Scaling morphometric pulmonary diffusing capacity to body mass: Wild and domestic mammals. *Respir Physiol* 44(1):61–86.
- Hoppeler H, Weibel ER (1998) Limits for oxygen and substrate transport in mammals. *J Exp Biol* 201(Pt 8):1051–1064.
- Post JR, Lee JA (1996) Metabolic ontogeny of teleost fishes. *Can J Fish Aquat Sci* 53(4): 910–923.
- Weibel ER, Knight BW (1964) A morphometric study on the thickness of the pulmonary air-blood barrier. *J Cell Biol* 21(3):367–396.
- Czopek J (1965) Quantitative studies on the morphology of respiratory surfaces in amphibians. *Acta Anat (Basel)* 62(2):296–323.
- Czopek J (1962) Vascularization of respiratory surfaces in some caudata. *Copeia* 3:576–587.
- Duncker H-R (2004) Vertebrate lungs: Structure, topography and mechanics. A comparative perspective of the progressive integration of respiratory system, locomotor apparatus and ontogenetic development. *Respir Physiol Neurobiol* 144(2–3):111–124.
- Sasaki N, Horinouchi H, Ushiyama A, Minamitani H (2012) A new method for measuring the oxygen diffusion constant and oxygen consumption rate of arteriolar walls. *Keio J Med* 61(2):57–65.
- Krogh A (1919) The rate of diffusion of gases through animal tissues, with some remarks on the coefficient of invasion. *J Physiol* 52(6):391–408.
- Huchzermeyer C, Berndt N, Holzhütter H-G, Kann O (2013) Oxygen consumption rates during three different neuronal activity states in the hippocampal CA3 network. *J Cereb Blood Flow Metab* 33(2):263–271.
- Uchida K, Matsuyama K, Tanaka K, Doi K (1992) Diffusion coefficient for O₂ in plasma and mitochondrial membranes of rat cardiomyocytes. *Respir Physiol* 90(3):351–362.
- Thao MT, Perez D, Dillon J, Gaillard ER (2014) Measuring the viscosity of whole bovine lens using a fiber optic oxygen sensing system. *Mol Vis* 20:125–131.
- Kreuzer F (1970) Facilitated diffusion of oxygen and its possible significance; a review. *Respir Physiol* 9(1):1–30.
- Kumosa LS, Routh TL, Lin JT, Lucisano JY, Gough DA (2014) Permeability of subcutaneous tissues surrounding long-term implants to oxygen. *Biomaterials* 35(29): 8287–8296.
- Dowse HB, Norton S, Sidell BD (2000) The estimation of the diffusion constant and solubility of O₂ in tissue using kinetics. *J Theor Biol* 207(4):531–541.
- Brafield A, Chapman G (1983) Diffusion of oxygen through the mesoglea of the sea anemone *Calliactis parasitica*. *J Exp Biol* 107(1):181–187.
- Chevillotte P (1973) Relation between the reaction cytochrome oxidase-oxygen and oxygen uptake in cells in vivo. The role of diffusion. *J Theor Biol* 39(2):277–295.
- Cheema U, et al. (2012) Oxygen diffusion through collagen scaffolds at defined densities: Implications for cell survival in tissue models. *J Tissue Eng Regen Med* 6(1):77–84.
- O'Loughlin MA, Whillans DW, Hunt JW (1980) A fluorescence approach to testing the diffusion of oxygen into mammalian cells. *Radiat Res* 84(3):477–495.
- van der Laarse WJ, des Tombe AL, van Beek-Harmsen BJ, Lee-de Groot MB, Jaspers RT (2005) Krogh's diffusion coefficient for oxygen in isolated *Xenopus* skeletal muscle fibers and rat myocardial trabeculae at maximum rates of oxygen consumption. *J Appl Physiol* (1985) 99(6):2173–2180.
- Rumsey WL, Schlosser C, Nuutinen EM, Robiolo M, Wilson DF (1990) Cellular energetics and the oxygen dependence of respiration in cardiac myocytes isolated from adult rat. *J Biol Chem* 265(26):15392–15402.
- Jones DP, Kennedy FG (1986) Analysis of intracellular oxygenation of isolated adult cardiac myocytes. *Am J Physiol* 250(3 Pt 1):C384–C390.
- Dejours P (1981) *Principles of Comparative Respiratory Physiology* (North-Holland, Amsterdam).
- White CR, Phillips NF, Seymour RS (2006) The scaling and temperature dependence of vertebrate metabolism. *Biol Lett* 2(1):125–127.
- Brown JH, Gillooly JF, Allen AP, Savage VM, West GB (2004) Towards a metabolic theory of ecology. *Ecology* 85(7):1771–1789.
- Grafen A (1989) The phylogenetic regression. *Philos Trans R Soc Lond B Biol Sci* 326(1233):119–157.
- Team RDC (2011) *R: A Language and Environment for Statistical Computing* (R Foundation for Statistical Computing, Vienna).
- Betancur-R R, et al. (2013) The tree of life and a new classification of bony fishes. *PLoS Curr* 5:1–33.
- Bininda-Emonds OR, et al. (2007) The delayed rise of present-day mammals. *Nature* 446(7135):507–512.
- Isaac NJ, Redding DW, Meredith HM, Safi K (2012) Phylogenetically-informed priorities for amphibian conservation. *PLoS One* 7(8):e43912.
- Jetz W, Thomas GH, Joy JB, Hartmann K, Moores AO (2012) The global diversity of birds in space and time. *Nature* 491(7424):444–448.
- Bergmann PJ, Irschick DJ (2012) Vertebral evolution and the diversification of squamate reptiles. *Evolution* 66(4):1044–1058.
- Pyron RA, et al. (2013) Genus-level phylogeny of snakes reveals the origins of species richness in Sri Lanka. *Mol Phylogenet Evol* 66(3):969–978.
- Jaffe AL, Slater GJ, Alfaro ME (2011) The evolution of island gigantism and body size variation in tortoises and turtles. *Biol Lett* 7(4):558–561.

59. Oaks JR (2011) A time-calibrated species tree of Crocodylia reveals a recent radiation of the true crocodiles. *Evolution* 65(11):3285–3297.
60. Rosindell J, Harmon LJ (2012) OneZoom: A fractal explorer for the tree of life. *PLoS Biol* 10(10):e1001406.
61. Baum BR (1992) Combining trees as a way of combining data sets for phylogenetic inference, and the desirability of combining gene trees. *Taxon* 41(1):3–10.
62. Ragan MA (1992) Phylogenetic inference based on matrix representation of trees. *Mol Phylogenet Evol* 1(1):53–58.
63. Creevey CJ, McInerney JO (2009) Trees from trees: Construction of phylogenetic supertrees using clann. *Methods Mol Biol* 537:139–161.
64. Goloboff PA, Farris JS, Nixon KC (2008) TNT, a free program for phylogenetic analysis. *Cladistics* 24(5):774–786.
65. Maddison W, Maddison D (2015) Mesquite: A modular system for evolutionary analysis. Version 2.75. 2011. Available at mesquiteproject.org.
66. Bininda-Emonds OR (2014) An introduction to supertree construction (and partitioned phylogenetic analyses) with a view toward the distinction between gene trees and species trees. *Modern Phylogenetic Comparative Methods and Their Application in Evolutionary Biology*, ed Garamszegi LZ (Springer, New York), pp 49–76.
67. Purvis A, Garland T, Jr (1993) Polytomies in comparative analyses of continuous characters. *Syst Biol* 42:569–575.
68. Tamura K, et al. (2012) Estimating divergence times in large molecular phylogenies. *Proc Natl Acad Sci USA* 109(47):19333–19338.
69. Paradis E, Claude J, Strimmer K (2004) APE: Analyses of phylogenetics and evolution in R language. *Bioinformatics* 20(2):289–290.
70. Felsenstein J (1985) Phylogenies and the comparative method. *Am Nat* 125(1):1–15.
71. Martins EP, Hansen TF (1997) Phylogenies and the comparative method: A general approach to incorporating phylogenetic information into the analysis of interspecific data. *Am Nat* 149(4):646–667.
72. Spatz H-C (1991) Circulation, metabolic rate, and body size in mammals. *J Comp Physiol B* 161(3):231–236.
73. Dlugosz EM, et al. (2013) Phylogenetic analysis of mammalian maximal oxygen consumption during exercise. *J Exp Biol* 216(Pt 24):4712–4721.
74. Taylor CR, Karas RH, Weibel ER, Hoppeler H (1987) Adaptive variation in the mammalian respiratory system in relation to energetic demand. *Respir Physiol* 69(1):1–127.
75. Hopkins SR (2007) Exercise induced arterial hypoxemia: The role of ventilation-perfusion inequality and pulmonary diffusion limitation. *Hypoxia and Exercise*, Advances in Experimental Medicine and Biology, eds Roach RC, Wagner PD, Hackett PH (Springer, New York), Vol 588, pp 17–30.
76. Weibel ER (1984) *The Pathway for Oxygen: Structure and Function in the Mammalian Respiratory System* (Harvard Univ Press, Cambridge, MA).
77. Gregory TR (2001) The bigger the C-value, the larger the cell: Genome size and red blood cell size in vertebrates. *Blood Cells Mol Dis* 27(5):830–843.
78. Hillman SS, Hancock TV, Hedrick MS (2013) A comparative meta-analysis of maximal aerobic metabolism of vertebrates: Implications for respiratory and cardiovascular limits to gas exchange. *J Comp Physiol B* 183(2):167–179.
79. Woods HA, Smith JN (2010) Universal model for water costs of gas exchange by animals and plants. *Proc Natl Acad Sci USA* 107(18):8469–8474.
80. Welch WR, Tracy CR (1977) Respiratory water loss: A predictive model. *J Theor Biol* 65(2):253–265.